

SAMPLE PREPARATION & SHIPPING INSTRUCTIONS

This document provides guidelines on how to prepare, quantify, and submit samples to Medikonia. Whether you are submitting DNA or RNA samples, it is essential that the appropriate instructions be followed to enable the successful completion of your project.

I. SAMPLE REQUIREMENTS

Sample quality directly impacts sequencing quality and subsequent bioinformatics analysis. Therefore, Medikonia has extensive sample quality control procedures to ensure submitted samples conform to requirements for downstream processing.

To guarantee the normal processing of your project, samples should meet the standards given below. If your samples do not meet these standards, or you are unable to produce higher-quality samples, please consult with your Medikonia Project Manager before shipping your samples.

Notes:

- 1) Input quantity should be determined by Qubit® instead of by NanoDropTM, and the final quantity and concentration should conform to Medikonia's specifications.
- 2) Samples not meeting these specifications should be designated as "at risk" by the customer, and will be subject to billing regardless of data quality. Please consult the Project Manager for further details.

1. Human Whole Genome/Exome Sequencing

		7				
Library	Comple Type	Amount (Qubit	Amount (Qubit®)			Purity
Type	Sample Type	Strongly Recommended	Required	Volume	Concentration	(NanoDrop TM /Agarose Gel)
Human	Genomic DNA	>2ua	> 1 mg	> 20I	> 20 ng/uI	OD260/280 = 1.8 - 2.0,
Whole	Genomic DNA	Genomic DNA $\geq 2 \mu g$ $\geq 1 \mu g$ $\geq 20 \mu L$ $\geq 20 ng$		\geq 20 ng/ μ L	no degradation, no contamination	
Genome/	Genomic DNA (PCR-free)	>2	> 1 mg	≥ 20 µL	> 20 ng/uI	OD260/280 = 1.8 - 2.0,
Exome	Genomic DNA (FCK-free)	≥ 2 µg	≥ 1 µg	\geq 20 μ L	$\geq 20 \text{ ng/}\mu\text{L}$	no degradation, no contamination
Sequencing	PCR products of	> 2	\ 1	> 20 I	> 20	Fragments should be longer than
/ Target	single-cell whole genome	≥ 2 µg	$\geq 1 \mu g$	\geq 20 μ L	$\geq 20 \text{ ng/}\mu\text{L}$	500 bp
Region	FFPE*	> 2	> 1.5 u.c.	-	-	Fragments should be longer than
Capture	FFPE"	≥ 3 µg	≥ 1.5 µg			1500 bp



* Formalin-fixed, paraffin-embedded

2. Plant & Animal Genome Sequencing

		Amount (Qub	oit®)			Purity (NanoDrop TM /Agarose Gel)	
Library Type	Sample Type	Strongly Recommended	Required	Volume	Concentration		
≤ 500 bp	Genomic DNA	$\geq 1.4~\mu g$	≥ 700 ng	\geq 20 μ L	$\geq 50 \text{ ng/}\mu\text{L}$		
Insert	Genomic DNA (PCR-free)	≥ 2 µg	≥ 1 µg	\geq 20 μ L	$\geq 20 \text{ ng/}\mu\text{L}$		
msert	Mitochondrion/Chloroplast DNA	$\geq 1.6 \ \mu g$	≥ 800 ng	$\geq 20~\mu L$	$\geq 50 \text{ ng/}\mu\text{L}$		
Genotyping by Sequencing	Genomic DNA	≥ 500 ng	≥ 300 ng	$\geq 10 \ \mu L$	$\geq 50 \text{ ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0, no degradation,	
2 Kb Insert	Genomic DNA	\geq 30 μ g	≥ 15 µg	\geq 20 μ L	$\geq 50 \text{ ng/}\mu\text{L}$	no contamination	
5 Kb Insert	Genomic DNA	\geq 30 μg	$\geq 15 \mu g$	$\geq 20 \ \mu L$	$\geq 50 \text{ ng/}\mu\text{L}$		
10 Kb Insert	Genomic DNA	≥ 50 µg	≥ 25 µg	\geq 20 μ L	$\geq 50 \text{ ng/}\mu\text{L}$		
> 10 Kb Insert	Genomic DNA	≥ 80 µg	≥ 40 µg	$\geq 20 \ \mu L$	$\geq 50 \text{ ng/}\mu\text{L}$		

3. Microbial Genome Sequencing

		Amount (Qubit®)				Purity
Library Type	ry Type Sample Type Strongly Required Required		Concentration	(NanoDrop TM /agarose gel)		
≤ 500 bp Insert Re	Genomic DNA	≥ 1.6 µg	≥ 800 ng	$\geq 20~\mu L$	$\geq 50 \text{ ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0, no degradation, no contamination
sequencing/ Meta Library	Genomic DNA (PCR-free)	≥ 2 µg	≥ 1 µg	$\geq 20~\mu L$	$\geq 50~\text{ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0, no degradation, no contamination
PCR-Free	Genomic DNA	≥ 10 µg	≥ 5 µg	$\geq 20~\mu L$	$\geq 50 \text{ ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0, no degradation, no contamination
Library for Amplicon	PCR Products*	≥ 400 ng	≥ 200 ng	$\geq 10 \ \mu L$	$\geq 20~\text{ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0, no degradation, no contamination



Genomic Services & Solutions

PCR Products**	≥ 300 ng	≥ 150 ng	≥ 10 μL	$\geq 20 \text{ ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0,
r CK Floducts	≥ 300 lig	≥ 130 lig	≥ 10 µL	≥ 20 Hg/μL	no degradation, no contamination

^{*} One PCR product for one library

4. Epigenetics Sequencing

			ıbit®)			Purity (NanoDrop TM / Agarose Gel)	
		Strongly Recommended	Required	Volume	Concentration		
Whole Genome	Genomic DNA (Genome size $\leq 1.5 \text{ G}$)	≥ 6 µg	≥3 µg	≥ 20 µL	$\geq 50 \text{ ng/}\mu\text{L}$	OD260/280 = 1.8 - 2.0,	
Bisulfite Sequencing	Genomic DNA (1.5G < Genome size ≤ 3.5 G)	≥ 12 µg	≥6 µg	≥ 20 µL	$\geq 50 \text{ ng/}\mu\text{L}$	no degradation, no contamination	
ChIP-Seq	ChIP-Seq DNA	≥ 100 ng	≥ 50 ng	≥ 10 µL	≥20 ng/μL	Main peak of 100 bp – 500 bp	

5. Transcriptome Sequencing

Library		Amount (Q	(ubit®)			RNA Integrity Number	Purity	
Library Type	Sample Type	Strongly Recommended	Required	Volume	Concentration	(Agilent 2100)	(NanoDrop TM)	
Eukaryotic	Total RNA (Animal)	≥ 2 µg	≥1 µg	\geq 20 μ L	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 6.8, smooth base line	OD260/280≥ 2.0,	
RNA-Seq	Total RNA (Plant and Fungus)	≥ 2 µg	≥ 1 µg	≥ 20 µL	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 6.3, smooth base line	$OD260/230 \ge 2.0$, no degradation,	
Prokaryotic RNA-Seq	Total RNA	≥ 6 µg	≥ 3 µg	\geq 20 μ L	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 6.0, smooth base line	no contamination	

6. Small RNA Sequencing

Library	Sample Type	Amount (Qubit®)	Volume	Concentration	RNA Integrity Number	Purity
---------	-------------	-----------------	--------	---------------	----------------------	--------

^{**} Multiple PCR products for one library (at least 2 different PCR products)



Genomic Services & Solutions

Type		Strongly Recommended	Required			(Agilent 2100)	(NanoDrop TM)
Eukaryotic	Total RNA (Animal)	≥ 6 µg	≥ 3 µg	$\geq 20~\mu L$	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 8, smooth base line	$OD260/280 \ge 2.0$, $OD260/230 \ge 2.0$,
small RNA Sequencing	Total RNA (Plant and Fungus)	≥ 6 µg	≥ 3 µg	≥ 20 µL	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 7.5, smooth base line	no degradation, no contamination

7. Long non-coding Sequencing

	Comple	Amount (Qubit®)				RNA Integrity Number	Purity	
Library Type	Sample Type	Strongly Recommended	Required	Volume	Concentration	(Agilent 2100)	(NanoDrop TM)	
Eukaryotic Long	Total RNA (Animal)	≥ 4 µg	≥ 2 µg	$\geq 20~\mu L$	$\geq 50 \text{ ng/}\mu\text{L}$	\geq 6.8, smooth base line	$OD260/280 \ge 2.0$,	
non-coding RNA Sequencing	Total RNA (Plant and Fungus)	≥ 4 µg	≥ 2 µg	≥ 20 µL	≥ 50 ng/µL	\geq 6.3, smooth base line	OD260/230 ≥ 2.0, no degradation, no contamination	

8. Pre-prepared library

Library volume requirement:

Data Amount	Volume Requirement*
< 30 G	≥ 10 µL
≥ 30 G	≥ 20 µL

^{*}High concentration samples should be diluted before delivery

(2) Library concentration: library concentration quantified by Qubit® 2.0 (Life Technologies): \geq 0.5 ng/uL



- (3) Insert size: dilute to 1 ng/µL before checking the insert size by Agilent 2100 Bioanalyzer.
- a) Insert size: insert + adapters (120 bp) \pm 50 bp (Does not apply to small RNA library)
- b) Main peak present, no multiple peaks, no adapter contamination and no primer dimers.

(4) Library concentration quantified by Q-PCR:

Platform	Concentration Requirement
HiSeq 2500	2 nM – 30 nM
MiSeq	4 nM – 30 nM
HiSeq X	3 nM – 30 nM

II. PRE-QUALITY CONTROL (QC) INSTRUCTIONS

Customers must provide the sample quality analysis results obtained using one of the following methods: Qubit®, NanoDropTM, agarose gel electrophoresis, or Agilent 2100. It is recommended samples be analyzed by Qubit/PicoGreen/gel electrophoresis (with quantity indicator), so that the results will correspond more closely to Medikonia QC results. NanoDropTM quantification is NOT recommended. If NanoDropTM is utilized for pre-QC quantification, Medikonia strongly recommends that you send more DNA/RNA for processing than the amounts given above.

For gel electrophoresis, the following conditions are recommended:

DNA: 1.0% agarose gel; 1.0% TAE solution; 100V for 40 min RNA: 1.0% agarose gel; 0.5× TBE solution; 180V for 16 min

Note:

Different electrophoresis conditions may generate a different, and potentially misleading, QC report on your samples. Therefore, it is highly recommended that



you adhere to the conditions recommended above for the initial check, and that you provide Medikonia with a picture of the gel.

III. DEMONSTRATIONS OF QUALIFIED DNA/RNA SAMPLES

1. Demonstration of Markers Used

Medikonia utilizes the following molecular size markers for sample quality control testing (Fig. 1).

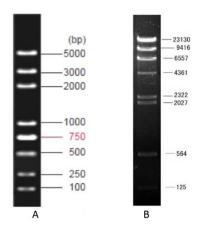


Fig. 1. (A) Trans $2K^{TM}$ Plus DNA Marker, (B) λ /HindIII DNA Marker, bp.

2. Demonstrations of DNA sample quality

2.1 Main types of sample quality



A qualified DNA sample is compared with common types of unqualified samples (Fig. 2):

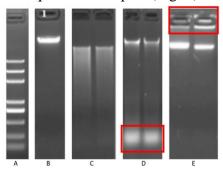


Fig. 2. Examples of DNA quality. (A) Trans2KTM Plus DNA Marker, (B) qualified sample, (C) degraded sample, (D) sample contaminated with RNA, (E) sample contaminated with protein. Red boxes denote areas of contamination.

2.2 Samples with

degradation

samples with

The gel picture illustrates degradation. Severe degradation can impact the quality of the prepared library and subsequent bioinformatics analysis (Fig. 3):

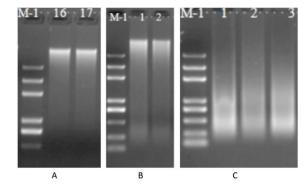


Fig. 3. DNA samples with degradation. Panels A, B, and C demonstrate increasing levels of DNA degradation. M-1, Trans2KTM Plus DNA Marker.



2.3 Samples with RNA contamination

RNA contamination of DNA samples (Fig. 4) can impede the library construction process. It is strongly recommended to digest your DNA samples with RNase before shipping.

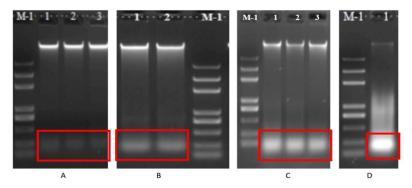


Fig. 4. DNA samples contaminated with RNA. Panels A - D demonstrate increasing levels of RNA degradation. Red boxes denote areas of contamination. M-1, Trans2 K^{TM} Plus DNA Marker.

2.4 Samples with protein

contamination

DNA samples can be contaminated by proteins, as illustrated in Fig. 5. It is recommended that you purify protein-contaminated DNA samples by affinity column. Please note that column purification will lead to some loss of DNA.



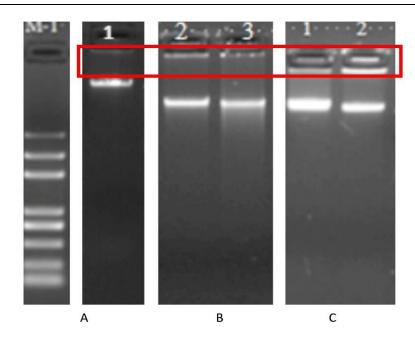


Fig. 5. DNA samples contaminated with protein. Panels A – C demonstrate increasing levels of protein contamination.

3. Demonstrations of RNA sample quality



3.1 Main types of sample quality

A qualified RNA sample is compared with common types of unqualified samples (Fig. 6):

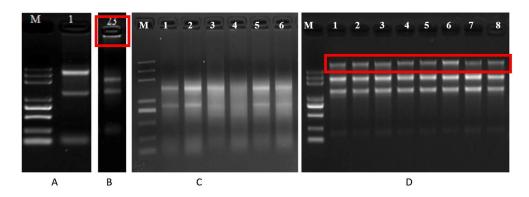


Fig. 6. Examples of RNA quality. (A) qualified sample, (B) sample with protein contamination, (C) samples with degratdation, (D) samples with genomic DNA contamination. Red boxes denote areas of contamination. M, Trans2KTM Plus DNA Marker.

3.2 Samples with protein contamination

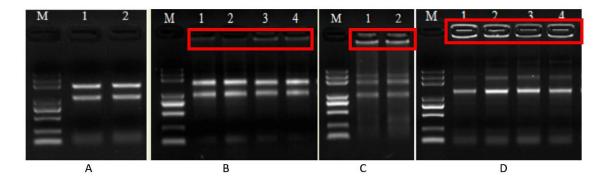


Fig. 7. RNA samples with protein contamination. Panels A – D demonstrate increasing levels of protein contamination. Red boxes denote areas of contamination. M, Trans2KTM Plus DNA Marker.



3.3 Agarose gel and Agilent 2100 analysis of RNA samples

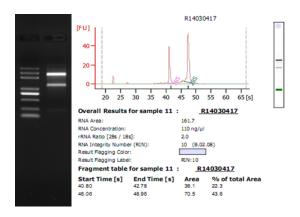


Fig. 8. An example of gel electrophoresis (left), and Agilent 2100 (right), results for an acceptable total RNA sample.

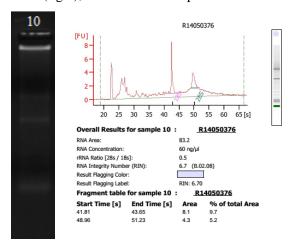


Fig. 10. An example of gel electrophoresis (left), and Agilent 2100 (right), results for an RNA sample with contamination.

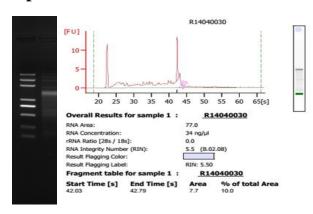


Fig. 9. An example of gel electrophoresis (left), and Agilent 2100 (right), results for a degraded total RNA sample.

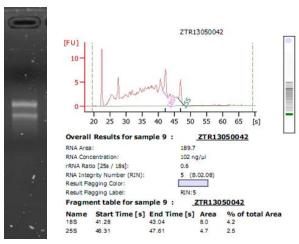


Fig. 11. An example of gel electrophoresis (left), and Agilent 2100 (right), results for a viscous total RNA sample.



IV. SAMPLE LABELING RECOMMENDATIONS

- 1. It is important to prevent the sample labels from being dissolved by solvents and from falling off the tubes. Use waterproof marker pen to write directly on the tube wall or lid is recommended strongly. Or you can also write the sample information on a paper/plastic label, stick the label onto the tube wall, and then secure the label to the tube by wrapping with clear, adhesive tape (e.g. Scotch tape) completely around the tube.
- 2. Please fill out and attach the Sample Information Form provided by Medikonia in the email before shipping the samples. Please make sure that the sample information on the Sample Information Form matches the labels on the tubes.

V. SAMPLE PACKING RECOMMENDATIONS

- 1. For DNA and RNA samples, Medikonia recommends 1.5 ml or 2 ml screw-cap DNase- and RNase-free microcentrifuge tubes. Please use Parafilm to seal each tube before packaging. Medikonia does not recommend shipping samples dissolved in organic solvents (such as absolute ethanol or isopropanol) because the solvents may cause leakage of the samples, which can result in cross-contamination between samples. If it is unavoidable to ship samples in organic solvents, please use screw-cap tubes and seal the opening of the tube with at least 10 layers of Parafilm.
- 2. In order to avoid crushing during shipping, Medikonia highly recommends placing the sample tubes in a container such as a 50-ml tube or a box with interior racks/holders. Cotton and absorbent papers can be used to prevent tubes from moving around inside the container.
- 3. RNA samples should be kept in dry ice during shipment. Genomic DNA samples should be kept in blue ice during shipment. Saliva samples should be shipped at room temperature.
- 4. In order to stick with our high quality control standards, 96-well plates and PCR stripe tubes are NOT acceptable containers for your sample shipping. The only container we allow for sample shipping is 1.5 ml or 2 ml tube. (See picture below).





Fig. 12. Recommended and prohibited tubes for sending samples

VI. COMPLETING THE SAMPLE SUBMISSION FORM

A Sample Information Form must be submitted for each sequencing service project. All information on the forms should be filled out carefully. Please submit the completed ELECTRONIC COPY via email to our local sales representative and enclose a HARD COPY in the shipment. In both copies, please make sure you mark the samples summary (Sample types and number) at the top of the Form (Fig. 13)

20 RNA Samples/40 DNA samples/50 Prepared Libraries Samples

RNA Sample Information Form

Notice:

- Completing this form with detail and accurate information will help us to serve you better (*fields are required to be filled).
- 2. Please enclose your samples with this sheet in hard copy and send a soft copy to Novogene representative.
- 3. If you have done Gel Electrophoresis Test, please attach the result below the form.



Fig. 13. Sample summary (top) and Sample Information Form

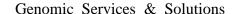
VII. SHIPPING SAMPLES TO MEDIKONIA

Disclaimer: The information below only constitutes a recommendation for shipping samples classified as "non-regulated materials" to our facility. At the time this document was prepared, gDNA/total RNA was not defined as a diagnostic specimen in the International Air Transport Association (IATA) packing instructions, and therefore no special packaging requirements are listed. Due to continuing changes in regulations, customers should always check with their safety office and/or shipping department to ensure regulatory compliance.

- 1. Ensure that all samples conform to our quality standards and that they are prepared and packaged according to the guidelines given above.
- 2. Please make sure to notify a Medikonia representative and to send the required documents before shipping your samples.
- 3. Select a reliable courier and choose the <u>priority option</u> for international shipments. Medikonia recommends FedEx (http://fedex.com/), UPS (www.ups.com), DHL (www.ups.com), TNT (www.ups.com), and World Courier (www.worldcourier.com). Whichever courier you choose, please make sure that the carrier can facilitate the importation of DNA or RNA samples, and dry ice packing (if applicable), into Hong Kong.
- 4. Sample transportation options:

	Lyophilize the DNA for shipping at ambient temperature
	Pack with ice packs/blue ice (2-8 °C)
DNA	Use the cold-chain transportation system (2-8 °C) of the courier
	DNA Stable (Liquid format, Biomatrica)
	Pack in dry ice (-60 °C – -80 °C)
	Lyophilize the RNA for shipping at 2-8 °C or ambient temperature
DNIA	Suspend RNA in 75% ethanol and ship on dry ice
RNA	RNAstable (Biomatrica)
	Pack in dry ice (-60 °C – -80 °C)

Note:





- 1) It is highly recommended that RNA samples be shipped in dry ice packaging. Other packaging/transportation methods may add impurities or cause slight degradation of the RNA.
- 2) The quantity of dry ice and ice bags needed varies with seasons (i.e., room temperature), transit time, and the thickness of Styrofoam box and receptacle. Please contact your local courier office for estimated transit time. Normally, dry ice is consumed (sublimates) at a rate of 5 kg per day.
- 5. Contact your local international courier and complete an INVOICE (commercial invoice, customs invoice, or pro-forma invoice) as required for customs, and include it with the shipment. Please complete the INVOICE as below:
 - 1) RNA or DNA Samples for Research Use Only
 - 2) Non-Dangerous, Non-Infectious
 - 3) No Commercial Value, Value for Customs Only
 - 4) Declare the value of the goods for customs [i.e. \$1.00 (USD) or €1.00 (EUR)]
 - 5) Number of samples and volumes [the # of samples, and the estimated volume]
 - 6) Type of container



Fig. 14. Courier INVOICE Example

Note: Please DO NOT include any other information about the source or about how you packed it. DO NOT include words such as "Human, Tissue, Cell, blood, blue ice, dry ice, etc." on the airway bill. Just write "RNA or DNA Samples for Research Use Only" (Please refer to the airway bill in the above picture). DO NOT write our company





name on the airway bill.

6. Package the samples with (1) a completed and detailed Sample Information Form; and (2) include any QC data for the samples if available (Qubit/Nanodrop/agarose gel electrophoresis/Agilent 2100). Pack the DNA and RNA samples according to the above options, and send the package to the address below: (Note: there is no zip code/postal code system in Hong Kong)

Email the Sample Information Form and Purchase Order (PO) to the Medikonia sales representative/project manager assigned to your project (indicated in the official quotation). Use the Sample Tracking Quote# xxx as the subject line in the email, and include the tracking information (courier name and tracking number) in the body of the email to help ensure that the samples arrive safely and without any delay.

7. After arriving at the Medikonia site, samples will be stored in -80 °C freezer. The Project Manager will be responsible for providing timely feedback to you on the progress of your project.

Qubit is a trademark of Life Technologies and Thermo Fisher Scientific.

NanoDrop is a trademark of NanoDrop Technologies LLC.

Agilent 2100 Bioanalyzer is a trademark of Agilent Technologies.

Trans2K Plus is a trademark of TransGen Biotech.